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Hemolymph Responses of The Xanthid Crab *Actaea hirsutissima* to Rhizocephalan *Sacculina ignorata* From Hurghada, Red Sea

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ABSTRACT

The effect of parasitic invasion by the Rhizocephala *Sacculina ignorata* on brachyuran crabs *Actaea hirsutissima* was studied. The organic constituents (glucose, total proteins, albumin, triglycerides and cholesterol) of the haemolymph of the infected and normal crab *Actaea hirsutissima* were estimated. A significant reduction in serum protein and albumin was observed in parasitized individuals; while serum glucose, cholesterol and triglycerides concentrations were significantly increased. Parasitism by *S. ignorata* dramatically affects the serum chemistry of infected crabs. T-test values were statistically highly significant between healthy and infected specimens.

INTRODUCTION

A common parasite of the Xanthid crab *Actaea hirsutissima* is the Rhizocephalan barnacle, *Sacculina ignorata* (Boschma, 1947b). This highly specialized cirriped consists of an extensive rootlet system termed (interna) that penetrates the haemocoel of the host. Histological analysis has demonstrated rootlet penetration of the host digestive system, allowing removal, sequestration and presumably utilization of host metabolites (Bresciani & Hoeg 2001). On maturation, the parasite forms an external egg sac (externa), which protrudes from the abdomen of chronically infected crabs. No host immune response to *S. carcini* has been observed in *Carcinus maenas*, and biological control methods are being actively studied in areas where the crab has been introduced artificially (Goddard *et al.*, 2005).

Stress responses occur in all animals when regulated physiological systems are extended beyond their normal range by external stressors. Failure of all or part of the integrated homeostatic response may lead to increasing physiological disturbance and ultimately death (Morris and Airriess, 1998). Indicators of such stress responses may therefore be useful in assessing the short-term well-being or long-term health status of an animal (Fossi *et al.*, 1997; Paterson and Spanoghe, 1997) and such indicators have received considerable attention in commercially important decapod crustacean species (Paterson and Spanoghe, 1997; Chang *et al.*, 1999). An important stressor is the infection of an animal by parasites (Thompson, 1983).

levels in their haemolymph (Krzynowek and Panunzio 1989; King *et al.*, 1990; O, Leary and Matthews 1990; Muriana *et al.*, 1993; Omar *et al.*, 1995). In pathogenic cases, Hudson (1995) studied the changes in biochemical parameters of haemolymph induced by parasites in crabs.

Overall, there has been scant information on haemolymph parameters of *A. hirsutissima* parasitized and healthy individuals. This study aims to evaluate the effect of parasitism on organic constituents (serum protein, albumin, cholesterol, triglycerides and glucose) of apparently uninfected crabs and infected individuals.

MATERIALS AND METHODS

Collection of crabs:

In the present study, adult crabs of *A. hirsutissima* were collected from Hurghada City (6.5 km North, beside Balm Beach Resort; Fig. 1), Red Sea (27°17'35.02" N and 33°45'37.68" E) during the period from February 2013 to June 2014. The specimens were collected by hand from the intertidal zone where they were hidden underneath small stones, coral rubbles and gravel. Subtidal inhabiting specimens were collected by snorkeling.

Invasion by macrobial or microbial agents causes changes in the haemolymph of host crabs. However, the measured parameters can increase, decrease or remain unchanged, depending on host or parasite species (Shirley *et al.*, 1986; Shields *et al.*, 2003). For example, a 23% decrease of total serum protein was observed in *Carcinus mediterraneus* infected by *Sacculina carcini*; while the same infection in *Pachygrapsus marmoratus* has a significant increase in these parameters. (Sanviti *et al.* 1981)

A number of researchers have suggested different methods for quantifying the stress reactions in crustaceans; these include the measurement of different hemocyte types in the hemolymph (Jussila *et al.*, 1997), physiological, biochemical (Paterson and Spanoghe, 1997; Stentiford *et al.*, 1999), molecular changes in tissue and hemolymph (Fossi *et al.*, 1997), and the elevated transcription of heat shock proteins during periods of stress (Chang *et al.*, 1999).

In crabs, studies on the levels of glucose, total proteins, total lipids, cholesterol, and albumin are still far from complete (Tsai *et al.*, 1984; Akpan, 1997). In contrast, these parameters were extensively studied in shrimps and recorded variable

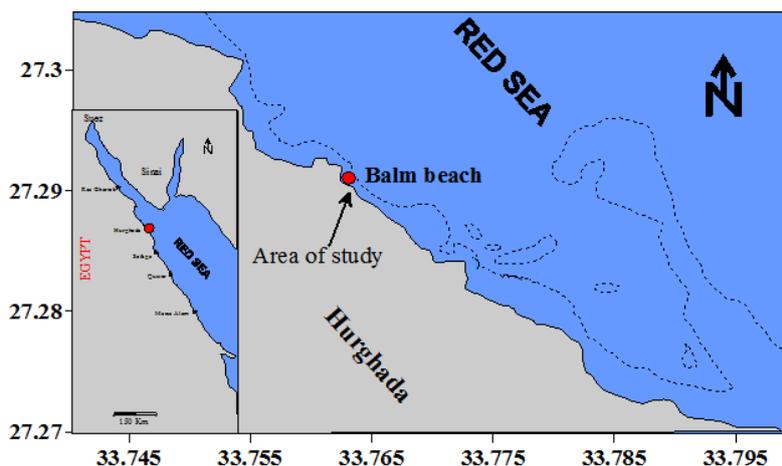


Fig. 1: A map showing the investigated site close to Balm Beach Resort. 6.5km North Hurghada, Red Sea, Egypt.

Parasite survey and identification:

For the parasite, it was identified according to Guerin-Ganivet (1911) and Boschma, (1947a, b; 1948).

Serum chemistry:

For biochemical analyses, 20 specimens of *A. hirsutissima* were collected,

sorted and measured to the nearest millimeter. The haemolymph was collected by heart puncture at the cardiac region of the carapace using a syringe. The collected haemolymph samples were put in marked vials, labeled, and then transported in ice – box to the laboratory, after that frozen till analysis.

At the laboratory, all the biochemical measurements were carried out using Shimadzu spectrophotometer (UV – Vis. 1201), Japan. Determination of organic constituent total proteins, albumin, glucose, cholesterol and triglycerides) were determined according to Hennery, 1968; Johnson *et al.*, 1999; Bergmeyer, 1974; Thomas, 1992; Cole *et al.*, 1997, respectively. All determinations were done

colorimetrically using Bicon Burbach kits Germany.

Data analysis: The data of these parameters were statistically analyzed using the student T- test.

RESULTS

Total proteins:

The values of total proteins in normal *A. hirsutissima* were higher than those infected specimens (Table 1) and were graphically represented in Figure (2). The higher values in normal crabs averaged, 5.80 ± 0.30 g/dl for females and 5.60 ± 0.39 g/dl for males with mean value (5.70 ± 0.23). These values slightly decreased to 4.03 ± 0.14 and 4.37 ± 0.31 g/dl in infected females and males, respectively, with mean value (4.20 ± 0.17).

Table 1: Mean values of (glucose, total proteins, albumin, triglycerides and cholesterol) in the haemolymph of *Actaea hirsutissima* Hurghada City from the Red Sea during the period from February 2013 to June 2014

Parameters		Total proteins (g\dl)	Albumin (g\dl)	Triglycerides (mg\dl)	Cholesterol (mg\dl)	Glucose (mg\dl)		
Crabs status								
normal	♂	X \pm SE	5.60 \pm 0.39	2.72 \pm 0.13	147.84 \pm 12.14	154.54 \pm 4.97	24.42 \pm 2.44	
		Range	4.80-6.97	2.29-3.03	112.90-172.40	145.70-173.70	17.70-29.90	
	♀	X \pm SE	5.80 \pm 0.30	3.09 \pm 3.27	152.40 \pm 24.90	174.30 \pm 7.99	30.20 \pm 6.67	
		Range	4.72-6.44	2.31-3.94	102.00-246.00	157.00-204.00	18.70-54.10	
	♂&♀		X \pm SE	5.70 \pm 0.23	2.91 \pm 0.16	150.12 \pm 13.08	164.42 \pm 5.53	27.31 \pm 3.49
	infected	♂	X \pm SE	4.37 \pm 0.31	2.02 \pm 0.12	195.62 \pm 18.28	220.66 \pm 16.02	49.48 \pm 1.63
Range			3.61-5.15	1.60-2.30	144.40-258.70	187.10-273.00	45.00-53.20	
♀		X \pm SE	4.03 \pm 0.14	1.90 \pm 0.27	231.34 \pm 13.11	210.60 \pm 4.03	38.40 \pm 2.05	
		Range	3.70-4.50	1.30-2.60	185.00-261.10	199.00-220.00	31.50-41.90	
♂&♀		X \pm SE	4.20 \pm 0.17	1.96 \pm 0.14	213.48 \pm 12.16	215.63 \pm 7.97	43.94 \pm 2.22	

The difference in protein levels was statistically highly significant between normal and parasitized crabs, as well as between normal and infected females. While the difference in protein levels were statistically significant between normal and infected males, as well as, between normal males and infected females (Table 2).

Albumin:

The haemolymph albumin of *A. hirsutissima* has the same pattern of protein for both normal and infected specimens of crabs. The albumin values were relatively higher in normal crabs than infected ones, (Table 1) and Figure (2). The albumin values were 3.09 ± 3.27 and 2.72 ± 0.13 g/dl for normal females and males, respectively, with

mean value (2.91 ± 0.16). Its values declined remarkably to 1.90 ± 0.27 g/dl and 2.02 ± 0.12 g/dl in infected females and males respectively with mean value averaged 1.96 ± 0.14 . The differences in albumin levels were statistically highly significant between normal and parasitized crabs, as well as between normal females and infected males. While the differences in albumin levels were statistically significant between normal and infected males. Also, between normal males and infected females (Table 2).

Cholesterol:

In contrast to total proteins and albumin, the cholesterol values for *A. hirsutissima* (Table 1 and Figure 2) were lower in normal crabs of both sexes than

infected ones. Cholesterol values were respectively, with mean value beings 154.54±4.97 mg/dl and 174.30±7.99 mg/dl for males and females of normal crabs, respectively, with mean value beings 164.42±5.53.

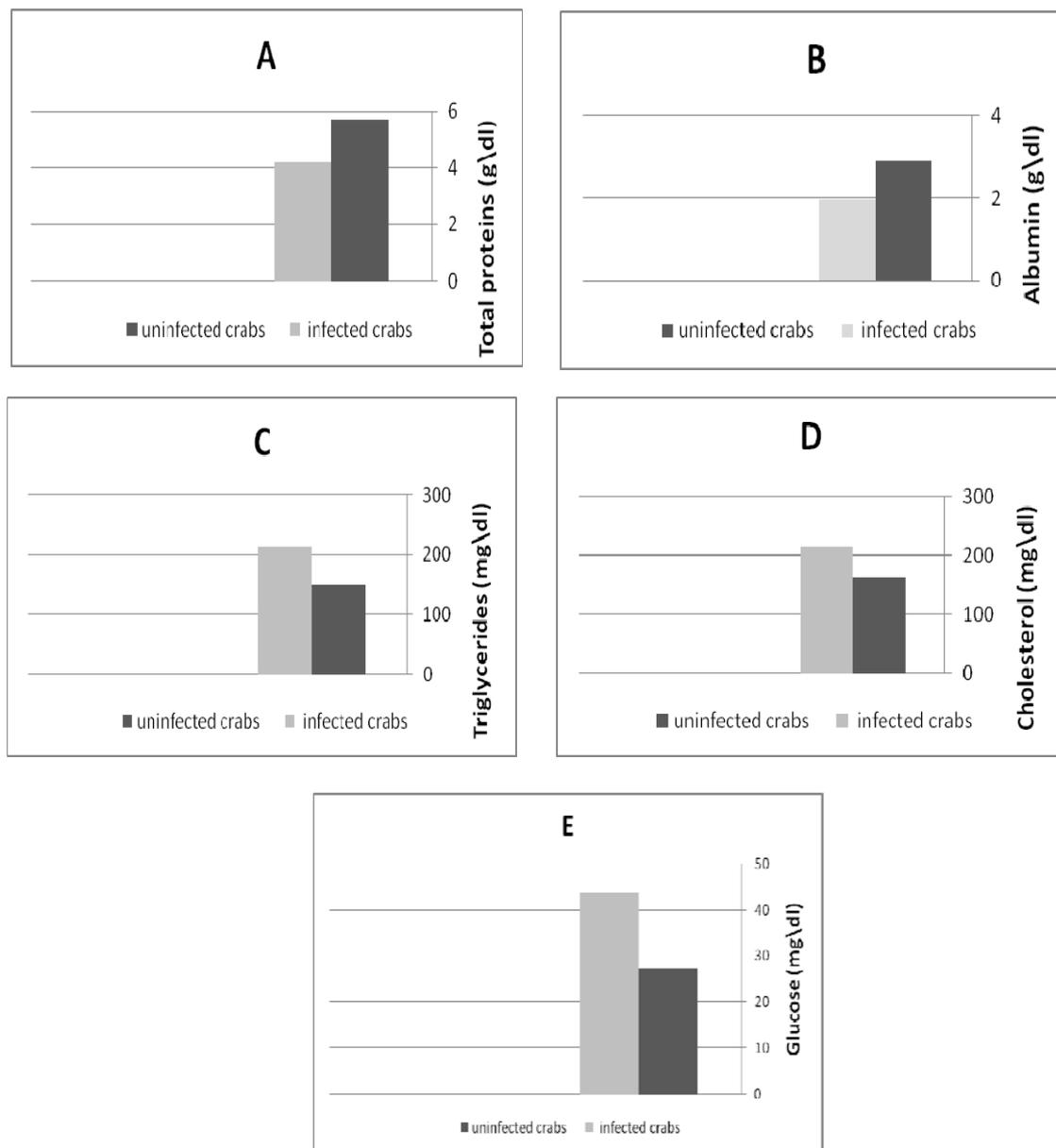


Fig. 2: Effect of *Sacculina ignorata* infection on *Actaea hirsutissima* organic constituents: (A) Total proteins (B) Albumin (C) Triglycerides (D) Cholesterol (E) Glucose.

The cholesterol levels are increased remarkably in infected crabs, to 210.60±4.03 mg/dl and 220.66±16.02 mg/dl for females and males respectively, with mean value beings 215.63±7.97. With only exception infected males and females, all statistical analyses were significant (Table 2).

Triglycerides:

The triglycerides values for *A. hirsutissima* (Table 1 and Figure 2) were

lower in normal crabs than infected ones, with lower values in males than females. It was 147.84±12.14 mg/dl and 152.40±24.90 mg/dl for males and females of normal crabs, respectively, with mean value beings 150.12±13.08. The triglycerides levels are increased remarkably to 231.34±29.32 mg/dl and 195.62±18.28 mg/dl for infected females and males respectively, with mean value being 213.48±12.16. The statistical analyses

(t-test) showed that the difference between males and infected females as well as healthy infected and normal females were significant, while those between normal and infected crabs were highly significant (Table 2).

Table 2: Students t-test values for organic constituents (glucose, total proteins, albumin, triglycerides and cholesterol) in the haemolymph of healthy and parasitized *Actaea hirsutissima* with *Sacculina ignorata*.

Crabs & sex	Total protein	Albumin	Triglycerides	Cholesterol	Glucose
Normal ♂ and ♀	0.449 NS	1.161NS	0.257NS	5.008**	0.972 NS
Normal and infected ♂	2.928*	3.373*	1.944 NS	5.470**	7.036 **
Normal ♂ and infected ♀	3.563*	3.420*	4.708**	6.775**	5.532**
Normal ♀ and infected ♂	2.641 NS	6.653**	1.269NS	3.719*	2.470 NS
Normal and infected ♀	4.168**	2.521 NS	3.349*	3.239*	1.144 NS
Infected ♂&♀	1.077 NS	0.342 NS	1.586 NS	0.519 NS	3,851*
Normal and infected crabs	5.070**	3.682**	3.755**	5.552**	3.535**

*: significant ($P < 0.05$).

** : highly significant ($P < 0.01$).

N.S: non significant ($P > 0.05$).

Glucose:

A remarkable increase in glucose level for males than females in infected crabs was noticed (Table and Figure 2). The glucose levels were 49.48 ± 1.63 and 38.40 ± 2.05 mg/dl for infected males and females, respectively, with mean value averaged 43.94 ± 2.22 . However, a reverse was noticed in normal ones. Their values were 24.42 ± 2.44 mg/dl in normal males and 30.20 ± 6.67 mg/dl in normal females with mean value was 27.31 ± 3.49 . The statistical analyses (t-test) showed that the difference between infected females and males were significant (Table 2), while those between normal males and infected males as well as normal males and infected females, in addition to healthy and parasitized crabs were highly significant (Table 2).

DISCUSSION

Crustaceans particularly those of commercial values, have high ratios of total protein than carbohydrates and total lipids (Warner, 1977; Amer *et al.*, 1991; Hashem, 1992; Abd El-Reheem, *et al.* 2003).

The current study found a significant reduction of serum protein and albumin in parasitized *A. hirsutissima*. This agrees with Powell and Rowley (2008) who found a significant reduction of serum protein in parasitized *Carcinus maenas*, infected with *Sacculina carcini* and also noticed that the

apparent loss of fibrillar protein from muscles. The parasite rootlet system is also capable of utilizing muscle-associated protein. Andrieux *et al.* (1980) observed the disappearance of a protein fraction in the haemolymph of *S. carcini*-parasitized crabs. However, the present results disagrees with Uglow (1969) who found that the total serum protein in control shore crabs was not significantly different to that of *S. carcini*-parasitized crabs, while earlier studies have suggested that parasitized individuals had significantly higher blood protein than uninfected counterparts (Drilhon, 1936).

Albumin is produced entirely in the liver and is of great importance in regulating the flow of water between the plasma and tissue fluid by its effect on colloid osmotic pressure as well as it is a negative acute phase protein and its concentration is decreased during inflammatory and parasitic infection (Mojabi, 2000, Stockham and Scott, 2002 and Kaneko, *et al.*, 2008). With an increase in load of gastrointestinal nematodes infection, the concentrations of total protein and albumin were decreased. (Chaichisemsari, *et al.*, 2011).

A drop in serum albumin level is usually associated with a decrease protein synthesis by the liver or increase protein loss through the gut or the kidney. Other possible cause of decrease in albumin may include malabsorption and increased protein need

secondary to infection (Halsted and Halsted, 1981; Cheesbrough, 1998). While El-Sayed *et al.* (2003) suggested that, the increase in the level of total proteins and albumin in the haemolymph of *Leptodius exaratus* individuals, infected with *Sacculina sp.* resulted from the destruction of both gonads and others tissues by the rootlets of the parasite within its host .

In Crustacea, the increase in haemolymph glucose levels is considered an important response of animals to stress (Jussila *et al.*, 1997; Hall and van Ham, 1998; Lorenzon, *et al.*, 2005). Hyperglycemia has been recorded following exposure of Crustacea to different stressors, such as emersion (Durand *et al.*, 2000), cold shock (Kuo and Yang, 1999), anoxia (Hall and van Ham, 1998) and pollutants (Lorenzon *et al.*, 2000) and has been observed previously in crabs infected by Rhizocephalan barnacles (Sanviti *et al.*, 1981; Shirley *et al.*, 1986).

In the current study, an increase in serum glucose recorded in the parasitized crabs. This agrees with Powell and Rowley (2008). They showed that an increase in serum glucose recorded in the parasitized crabs could result from the translocation of glucose from either hepatopancreas or muscle tissues. But they noticed that, no decrease of hepatopancreatic glycogen was observed in infected animals. So, they suggest that glucose may be mobilized from other tissues that store glycogen which were not assayed in their study. Muscle glycogen content was significantly depleted in *Nephrops norvegicus* infected with the dinoflagellate *Hematodinium sp.* (Stentiford *et al.*, 2000).

Change in carbohydrate dynamics in infected crustaceans can also result from alterations in the endocrine system. Stentiford *et al.* (2001) concluded that the release of glucose into the haemolymph is mediated by the crustacean hyperglycemic hormone through the mobilisation of intracellular glycogen stores. Whereas, they found that *N. norvegicus* infected with *Hematodinium sp.* suffered depleted glucose and hepatopancreatic glycogen reserves, as

infection disrupted the feedback loop for crustacean hyperglycemic hormone. In contradictory with the present result, El-Sayed *et al.*, (2003) reported that the sharp decline in infected specimens of *L. exaratus* infected with *Sacculina sp.* indicate sharp depletion in haemolymph glucose which may be consumed by the parasites.

The total lipid is generally low in crustaceans (Warner, 1977; Amer *et al.*, 1991; Hashem, 1992; El-Zawahry *et al.*, 1997; Abd El-Reheem, *et al.* 2003). Cholesterol is one of the most important constituents of lipids in haemolymph of crustaceans and is the basic constituent of sex hormones (Highnam & Hill, 1979). During this study, the cholesterol level was high in the haemolymph of the parasitized crabs than healthy ones. El-Sayed *et al.*, (2003) mentioned that remarkable changes were evident and closely correlated with the morphological (moulting) and physiological (reproductive) changes of the crabs of the two sexes for two different species. A remarkable decrease was recorded in infected males and females of *L. exaratus* as a result of infection with *Sacculina sp.* (El-Sayed *et al.*, 2003). As shown herein, infected crabs have high circulating levels of triglycerides and cholesterol. These results are in agreement with those published by Robson (1911) who explain the effect of the Rhizocephalan parasite *Sacculina neglecta* upon the sexual physiology of its host, the spider-crab, *Inachus mauritanicus*. He reported that the increased fat in the blood of the infected crabs, *Inachus mauritanicus* parasitized with *Sacculina neglecta* is the result of an increased activating of fat that would otherwise have been stored up or is due to an increased initial supply.

In the light of the previous facts, it is obvious that parasitism by *Sacculina ignorata* dramatically affects the serum chemistry of infected crab *A. hirsutissima*. Whereas, it causes a significant reduction in serum protein and albumin and increases serum glucose, cholesterol and triglycerides concentrations in parasitized individuals.

REFERENCES

- Abd El-Reheem, A. M. A., El-Zawhry, E. I. A. and Mohamed, E.F. (2003). Comparative study of some physiological parameters in the haemolymph of two crabs, *Ocypoda saratan* and *Grapsus albolineatus* from protected areas of Red Sea, Sc. J. Az. Med. Fac. (Girls), 24 (2):1545-1555.
- Akpan, E.J. (1997). Proximate composition of edible blue crab, *Callinectes sapidus*. J. Food Sci. & Tech. Mysore, 34 (1): 59-60.
- Amer, H.A., Sedik, M.F., Khalafalla, F.A. and Awad, H.A.E.G. (1991). Results of chemical analysis of prawn muscle as influenced by sex variation, Nahrung., 35(2): 133-138.
- Andrieux, N., Herberts, C., Frescheville, J.D. (1980) Protein metabolism and parasitism in the crab *Carcinus maenas*-effects of *Sacculina carcini* (Crustacea, Rhizocephala) on serum and epidermal proteins of host. Can J. Zool., 58: 580-585.
- Bergmeyer, H.V. (1974): Methods of enzymatic analysis, verlage chemie. Associated Press, New Yourk, P. 1196.
- Boschma, H. (1947a). Three successive layers of external cuticle in *Sacculina leptodidae*. Proc. Kon. Ned. Akad. Wetensch. Amsterdam, 50:3-9.
- Boschma, H. (1947b). The rhizocephalan parasites of the crab *Actaea hirsutissima* (Rüpp.). Proc. Kon. Ned. Akad. Wetensch. Amsterdam, 50: 272 -278.
- Boschma, H. (1948). *Sacculina leptodidae* Guér.-Gan., a Parasite of three different crabs. Zool. Meded. Mus. Leiden, 30: 49 - 71.
- Bresciani, J. and Hoeg, J. T. (2001). Comparative ultrastructure of the root system in rhizocephalan barnacles (Crustacea: Cirrhopedia: Rhizocephala). J. Morphol, 249:9-42.
- Chaichisemsari, M., Eshratkhan, B., Maherisis, N., Sadaghian, M. and Hassanpour, S. (2011). Evaluation of total protein, albumin, globulin and blood urea nitrogen concentrations in gastrointestinal nematodes infected sheep. Global Veterinary, 6(5): 433-437.
- Chang, E. S., Chang, S. A., Keller, R., Reddy, P. S., Snyder, M. J. and Spees, J. L. (1999). Quantification of stress in lobsters: Crustacean hyperglycemic hormone, stress proteins and gene expression. Am. Zool., 39: 487-495.
- Cheesbrough, M. (1998). District laboratory practice in tropical countries, Part 1. Cambridge University Press, Cambridge. pp. 355-358.
- Cole, T. G., Klotzsch, S. G. and McNamara, J. (1997). Measurement of triglyceride concentration. In: Rifai, N.; Warnick, G. R. and Dominiczak, M. H., (Eds.), Handbook of lipoprotein testing. Washington: AACC Press, pp. 115-126.
- Drilhon, A. (1936) Quelques constants chimiques et physicochimiques du milieu interieur du crabe sacculine (*Carcinus maenas*). CR Hebdomaires Séances Acad Sci., 202:981-982
- Durand, F., Devillers, N., Lallier, F.H. and Regnault, M. (2000). Nitrogen excretion and change in blood components during emersion of the subtidal spider crab *Maia squinado* (L.). Comp. Biochem. Physiol., 127A: 259-271.
- El-Sayed, A. A. M., El-Damhougy, K. A., Saber, S. A. A. and Fouda, M. M. A. (2003). Mobilization of haemolymph constituents during moulting stages of the intertidal crabs, *Leptodius exaratus* and *Metopograpsus messor* (Brachyura, Crustacea) from A' in Sukhna, Suez Gulf, Egypt. J. Aquat. Biol. and Fish., 7(1):141-158.
- El-Zawahry, E.I.A., El-Sayed, A.A.M. and Zaaqouk S.A. (1997). Some biochemical parameters of haemolymph of Xanthid crab, *Leptodius exaratus* and Portunid crab *Thalamita poissoni* from the Suez Gulf. Egypt. J. Aquat. Biol. and fish., 1(1):17-25.
- Fossi, M. C., Savelli, C., Casini, S., Franchi, E., Mattei, N. and Corsi, I. (1997). Multi-response biomarker approach in the crab *Carcinus aestuarii* experimentally exposed to benzo (a) pyrene,

- polychlorobiphenyls and methyl-mercury. *Biomarkers*, 2: 311–319.
- Goddard, J.H.R., Torchin, M.E., Kuris, A.M. and Lafferty, K.D. (2005). Host specificity of *Sacculina carcini*, a potential biological control agent of introduced European green crab *Carcinus maenas* in California. *Biol Invasions*, 7: 895–912.
- Guerin-Ganivet, J. (1911). Contribution a Petude systematique et biologique des Rhizocephales. *Trav. Scient. Lab. Zool. et Physiol. Marit. Concarneau*, 3: 1–97.
- Hall, M. R., and van Ham, E. H. (1998). The effects of different type of stress on blood glucose in the giant tiger prawn *Penaeus monodon*. *J. World Aquacult. Soc.*, 29: 290–299.
- Halsted, J.A. and Halsted, C.H. (1981). The laboratory in clinical medicine: Interpretation and application, 2nd edition. WB. Saunders Company, Philadelphia., pp. 281-283.
- Hashem, H.O. (1992). Observations on the biochemical changes in some organs of *Portunus pelagicus* (L.) in relation to maturation. *J. Egypt. Ger. Soc. Zool.*, 7 (A), Comparative physiology, 441-452.
- Hennery R.J. (1968). *Chinical Chemistry principles and Technics* Haper and Row. New York P.197.
- Highnam, K.C. and Hill, L. (1979). Endocrine mechanism in Crustacea I in: the compartive endocrinology of the invertabrates. Barrington, E.J.W and Willis, A.J. eds, the English language Book society and Arnold, E. (publishers) Ltd. pp. 209-224.
- Hudson, D.A. (1995). Biochemical parameters of the serum of the sand crab, *Portunus pelagicus*, with reference to the parasitic dinoflagellate *Hematodinium australis*. *Bull. Eur. Assoc. Fish pathol.*, 15(6):202-205.
- Johnson, A.M., Rohlf, E.M. and Silverman, L.M. (1999). Proteins, p 477. In Burtis CA, Ashwood ER (eds), *Tietz textbook of clinical chemistry*, ed 2. WB Saunders, Philadelphia, PA.
- Jussila, J., Jago, J., Tsuetnenko, E., Dunstan, B. and Evans, L. H. (1997). Total and differential haemocyte counts in western rock lobsters (*Panulirus cygnus* George) under post-harvest stress. *Mar. Freshw. Res.*, 48: 863–867.
- Kaneko, J.J., Harvey, J.W. and Bruss, M.L. (2008). *Clinical biochemistry of domestic animal*, 6th edn, Academic Press, USA, pp: 117-156.
- King, I., Childs, M.T., Dorsett, C., Ostrander, J.G. and Monsen, E.R. (1990). Shell fish; proximate composition, minerals, fatty acids and sterols. *J. Amer. Dietetic Association*, 90(5):677-685.
- Krzynowek, J. and panunzio, L.J. (1989). Cholesterol and fatty acids in several species of shrimp. *J. Food Sci.*, 54 (2):237-239.
- Kuo, C.M. and Yang, Y.H. (1999). Hyperglycemic responses to cold shock in the freshwater giant prawn, *Macrobrachium rosenbergii*. *J. Comp. Physiol.*, 169(B): 49-54.
- Lorenzon, S., Francese, M. and Ferrero, E.A. (2000). Heavy metal toxicity and differential effects on the hyperglycemic stress response in the shrimp *Palaemon elegans*. *Arch. Environ. Contam. Toxicol.*, 39: 167-176.
- Lorenzon, S., Edomi, P., Giulianini, P.G., Mettullo, R. and Ferrero, E. A. (2005). Role of biogenic amines and CHH in the crustacean hyperglycemic stress response. *J. Exp. Biol.* 208: 3341-3347; doi: 10.1242/jeb.01761.
- Mojabi, A. (2000). *Veterinary clinical biochemistry (in Farsi)*, 2nd ed. Noorbakhsh Press, Tehran, 149: 444-453.
- Morris, S. and Airriess, C. N. (1998). Integration of physiological responses of crustaceans to environmental challenge. *S. African J. Zool.* 33, 87–106.
- Muriana, F.J.G., Ruiz-Gutierrez, V. and Bolufer, J. (1993). Phospholipid fatty acid composition of hepatopancreas and muscle from the prawn, *Panaeus japonicus*. *J. Biochemistry –Tokyo.* 114 (3): 404-407.

- O'leary, C.D. and Matthews, A.D. (1990). Lipid class distribution and Fatty acids composition of wild and farmed prawn, *Panaeus monodon* (Fabricius), Aquacult., 89(1):65-81.
- Omar, M.A.K., Malik, O.A. and Ismail, N. (1995). Cholesterol content of some Malaysian marine prawn species. Asean-food Jor., 10 (1):39-40.
- Paterson, B. D. and Spanoghe, P. T. (1997). Stress indicators in marine decapod crustaceans, with particular reference to the grading of western rock lobsters (*Panulirus cygnus*) during commercial handling. Mar. Freshw. Res., 48:829-834.
- Powell, A. and Rowley, A. F. (2008). Tissue changes in the shore crab *Carcinus maenas* as a result of infection by the parasitic barnacle *Sacculina carcini*. Dis Aquat Org., 80: 75–79.
- Robson, G. C. (1911). The Effect of *Sacculina* upon the fat metabolism of its host. Q.J.M.S. 57, pp. 267-278.
- Sanviti, G., Romestand, B. and Trilles, J.P. (1981). Les sacculines (*Sacculina carcini* Thompson, 1836) de *Carcinus mediterraneus* et *Pachygrasus marmoratus*: comparaison immunochemique: étude comparée de leur influence sur la composition protéique de l'hémolymph des deux hôtes. Parasitol Res 64:243–251.
- Shields, J.D., Scanlon, C. and Volety, A. (2003). Aspects of the pathobiology of the blue crabs, *Callinectes sapidus*, infected with the parasitic dinoflagellate *Hematodinium perezii*. Bull. Mar. Sci., 72: 519–535.
- Shirley, S.M., Shirley, T.C. and Meyers, T.R. (1986). Haemolymph responses of Alaskan king crabs to rhizocephalan parasitism. Can J Zool 64:1774–1781.
- Stentiford, G. D., Neil, D. M. and Coombs, G. H. (1999). Changes in the plasma free amino acid profile of the Norway lobster, *Nephrops norvegicus*, at different stages of infection by a parasitic dinoflagellate (genus *Hematodinium*) Diseases Aquat. Organ. 38, 151–157.
- Stentiford, G.D., Neil, D.M. and Coombs, G.H. (2000). Alterations in the biochemistry and ultrastructure of the deep abdominal flexor muscle of the Norway lobster *Nephrops norvegicus* during infection by a parasitic dinoflagellate of the genus *Hematodinium*. Dis Aquat Org 42:133–141.
- Stentiford, G. D., Chang, E. S., Chang, S.A. and Neil, D.M. (2001). Carbohydrate dynamics and the crustacean hyperglycaemic hormone (CHH): effects of parasitic infection in Norway lobsters (*Nephrops norvegicus*). Gen Comp Endocrinol, 121:13–22.
- Stockham, S.L and Scott, M.A. (2002). Fundamental of veterinary clinical pathology. Iowa state Press, USA, pp: 540-546.
- Tasi, D. E., Chen, H. C. and Tsai, C. F. (1984). Total lipid and cholesterol content in the blue crab, *Callinectes sapidus* Rathbun. Comparative-Bioch. and phys., B, 78(1):27-31.
- Thomas L. (1992). Enzymatic colorimetric method to determine the cholesterol. Lab. And Diagnose 4ed. 208: 964-980
- Thompson, S. N. (1983). Biochemical and physiological effects of metazoan endoparasites on their host species. Comp. Biochem. Physiol. B 74: 183–211.
- Uglow RF (1969) Haemolymph protein concentrations in portunid crabs – III. The effect of *Sacculina*. Comp Biochem Physiol 31: 969–973
- Warner, G. F. (1977). Association. In: The Biology of crabs. pp. 81-84, Elek Science, London.